



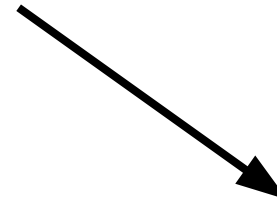
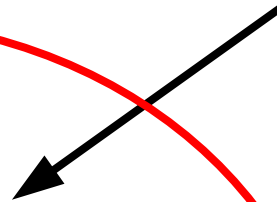
Mutagenesis & in-vitro evolution

The course schedule is
subject to modifications!



Mutagenesis & in-vitro evolution

Mutagenesis

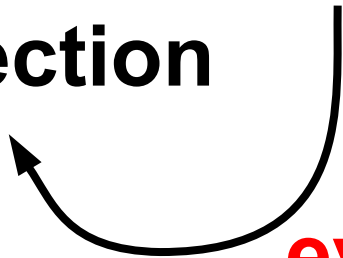
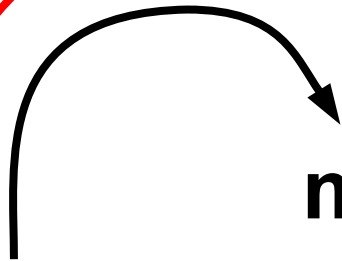


**Random
mutagenesis**

**Site-directed
mutagenesis (SDM)**

Selection

**in-vitro/
directed
evolution**





Site-directed mutagenesis & in-vitro evolution

Types of mutations you may want to introduce:

- Point mutations
- Deletions
- Insertions

Technically, cloning an insert into a vector is equivalent to insertional mutagenesis (“cassette mutagenesis”)



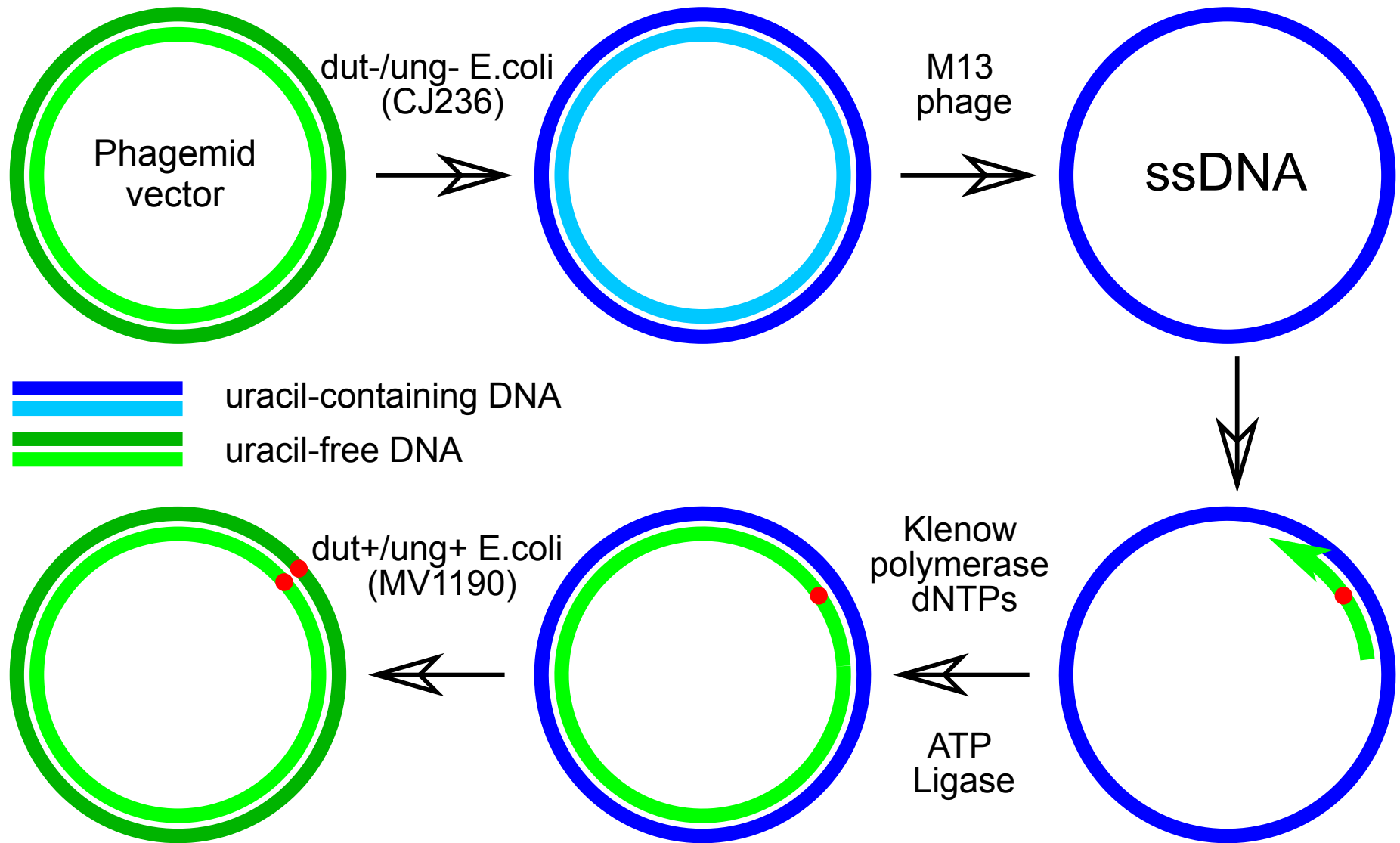
Site-directed mutagenesis & in-vitro evolution

Methods

- Kunkel-Method, MutaGene[®] (Bio-Rad)
- Whole plasmid mutagenesis/Inverse PCR mutagenesis: QuikChange[®] (Agilent), Q5[®] SDM Kit (NEB), Phusion[®] SDM Kit (ThermoScientific)
- Insert PCR mutagenesis: Simple and overlap extension mutagenesis
- SDM by Gibson-Assembly or In-Fusion



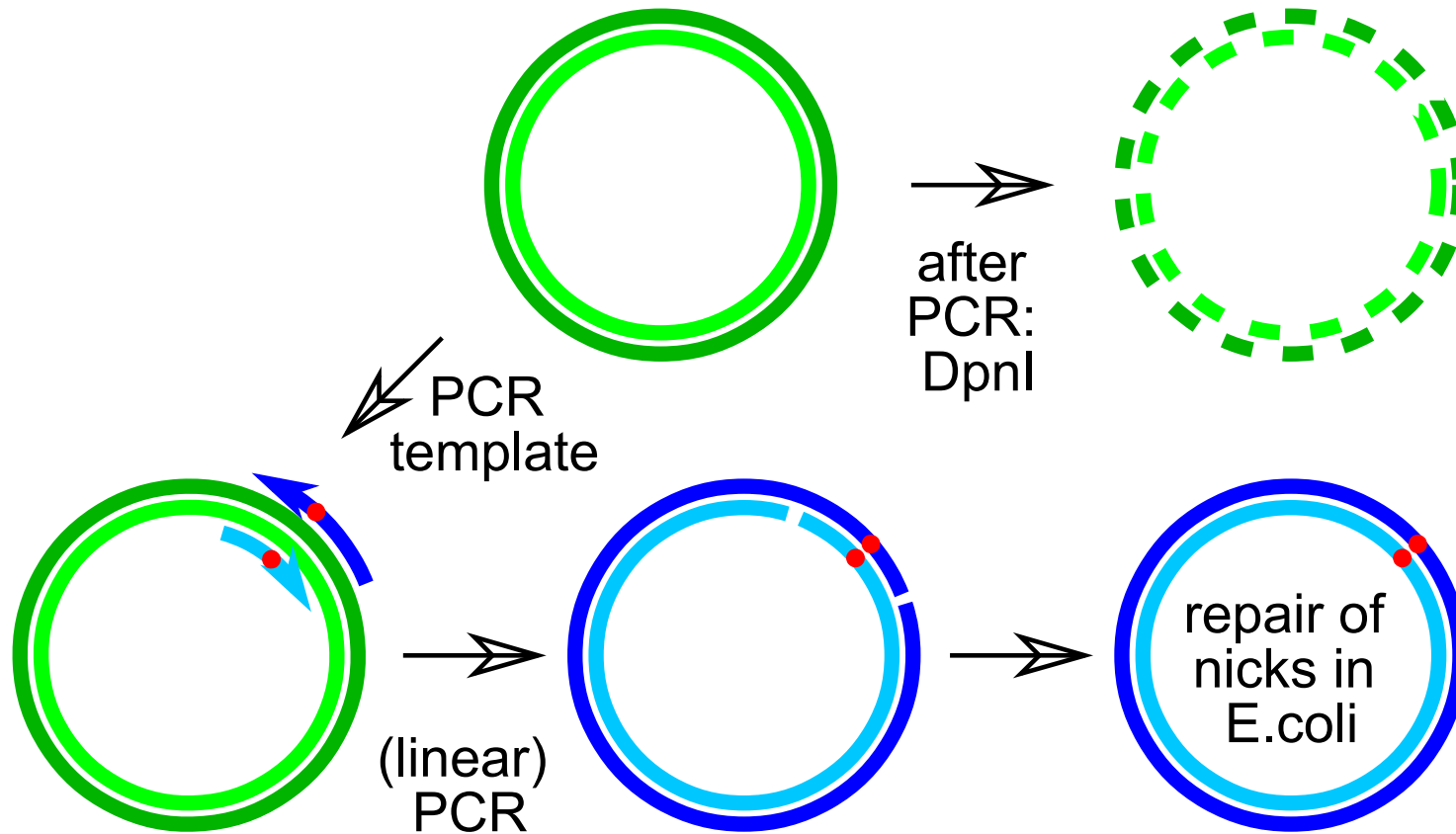
Kunkel method



ung- = uracil N-glycosylase-deficient => high intracellular levels of dUTP
dut- = dUTPase-deficient => prevents removal of uracil from DNA



QuikChange

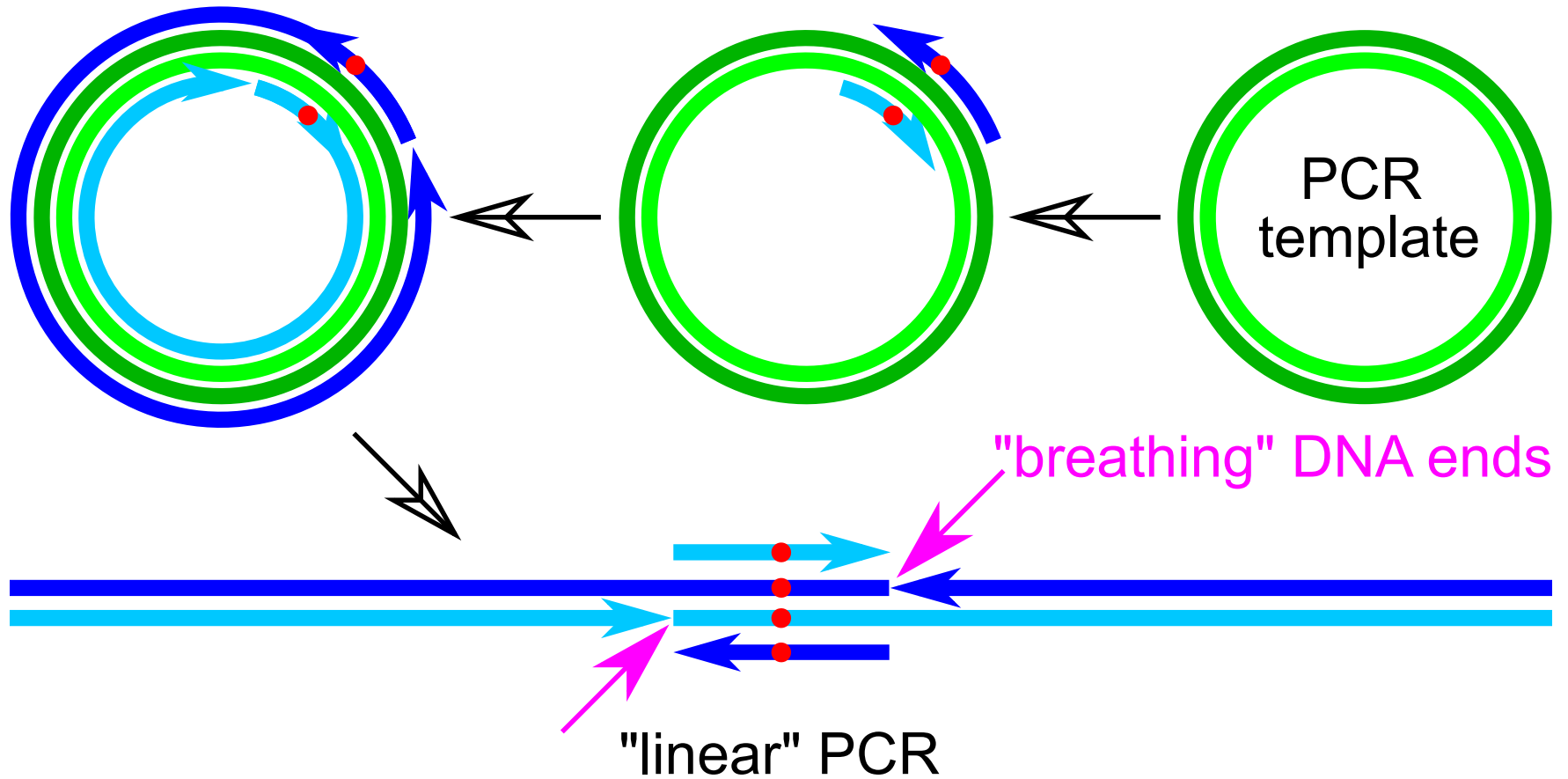


Limitations

- **PCR efficiency**
 - Primer dimers
 - Length of construct
 - DNA secondary structure (very high/low GC content)
- **Type of mutation**
 - Point mutations
 - Small insertions & deletions

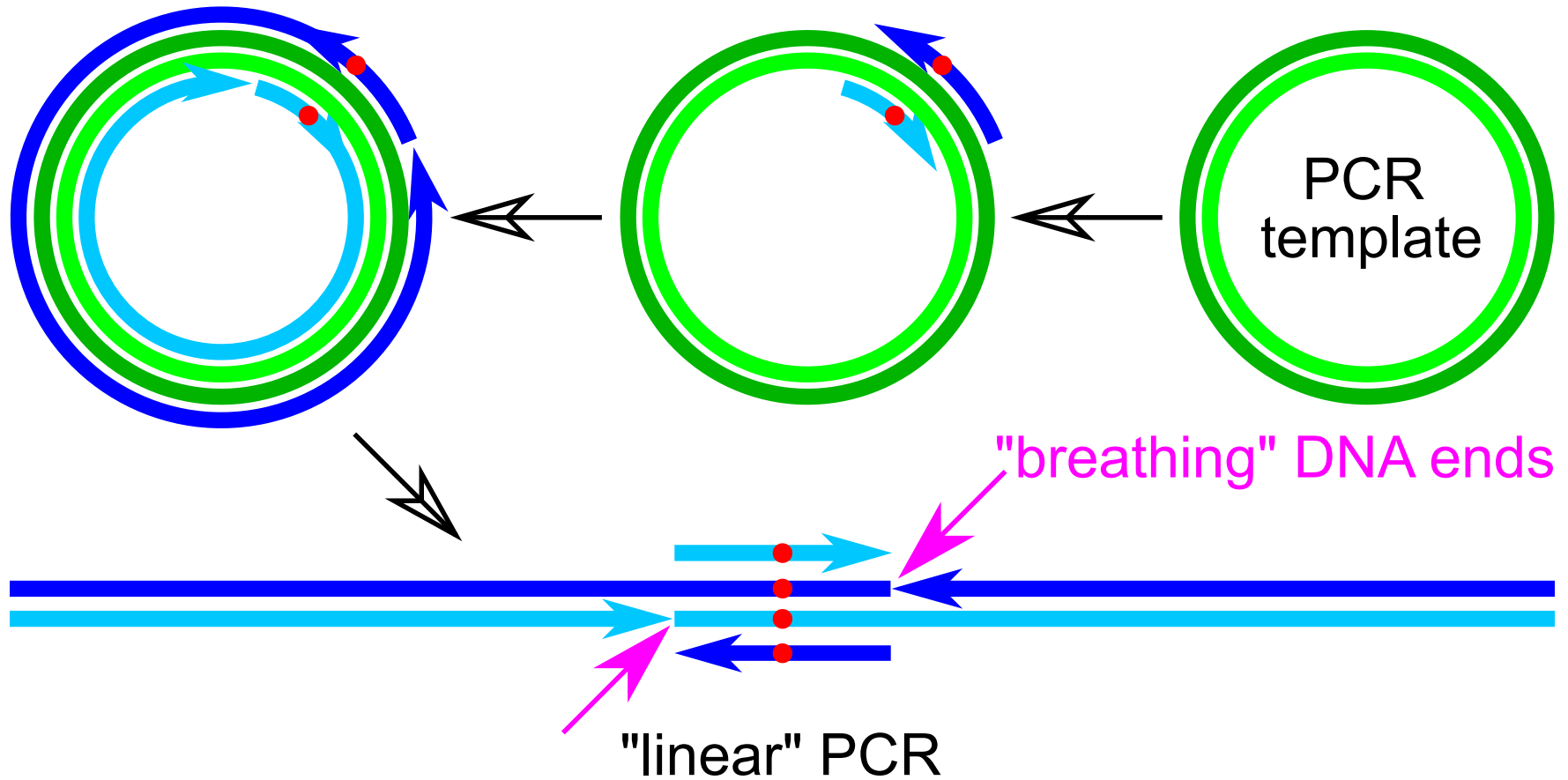


QuikChange



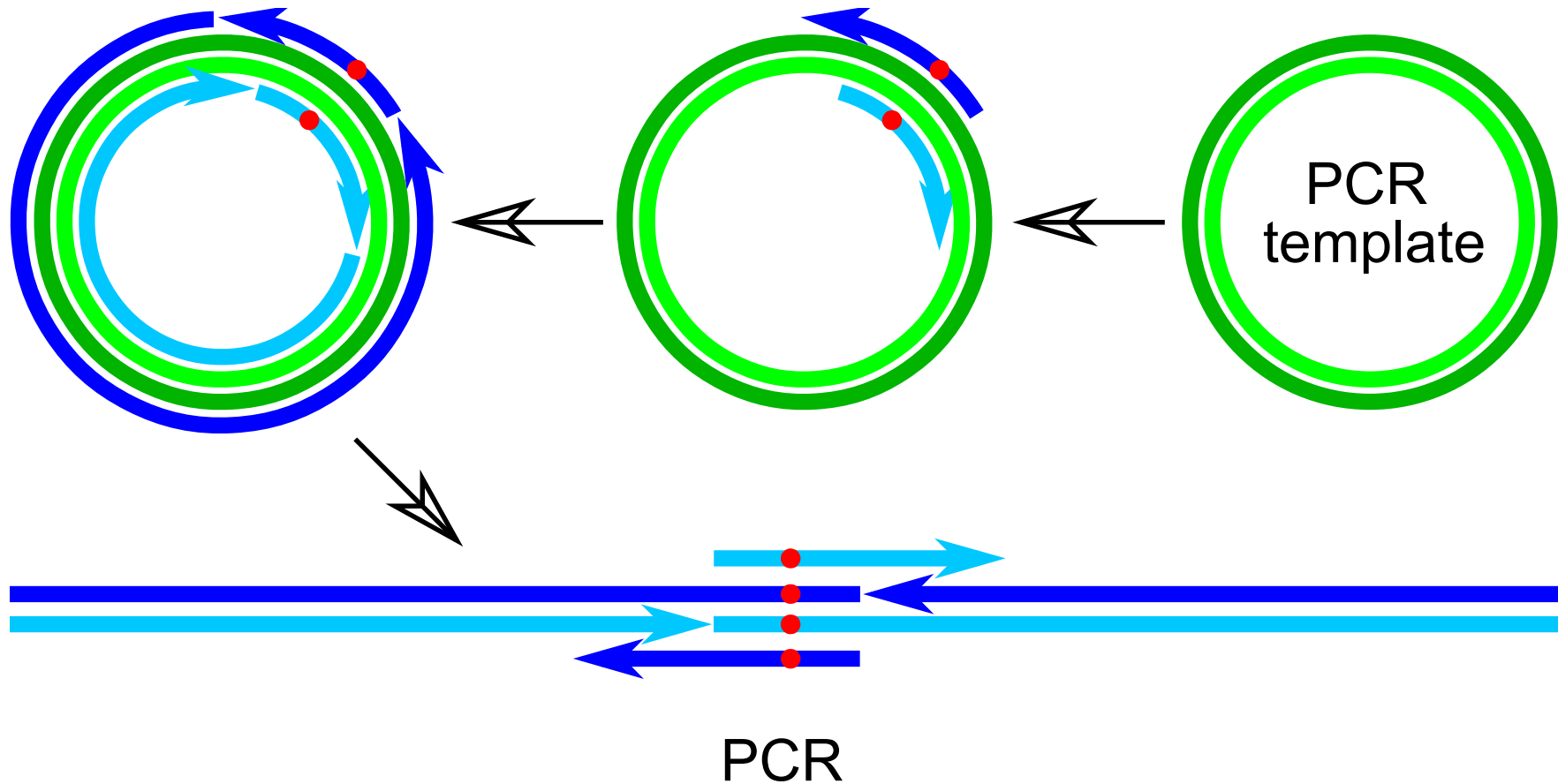


Improved QuikChange



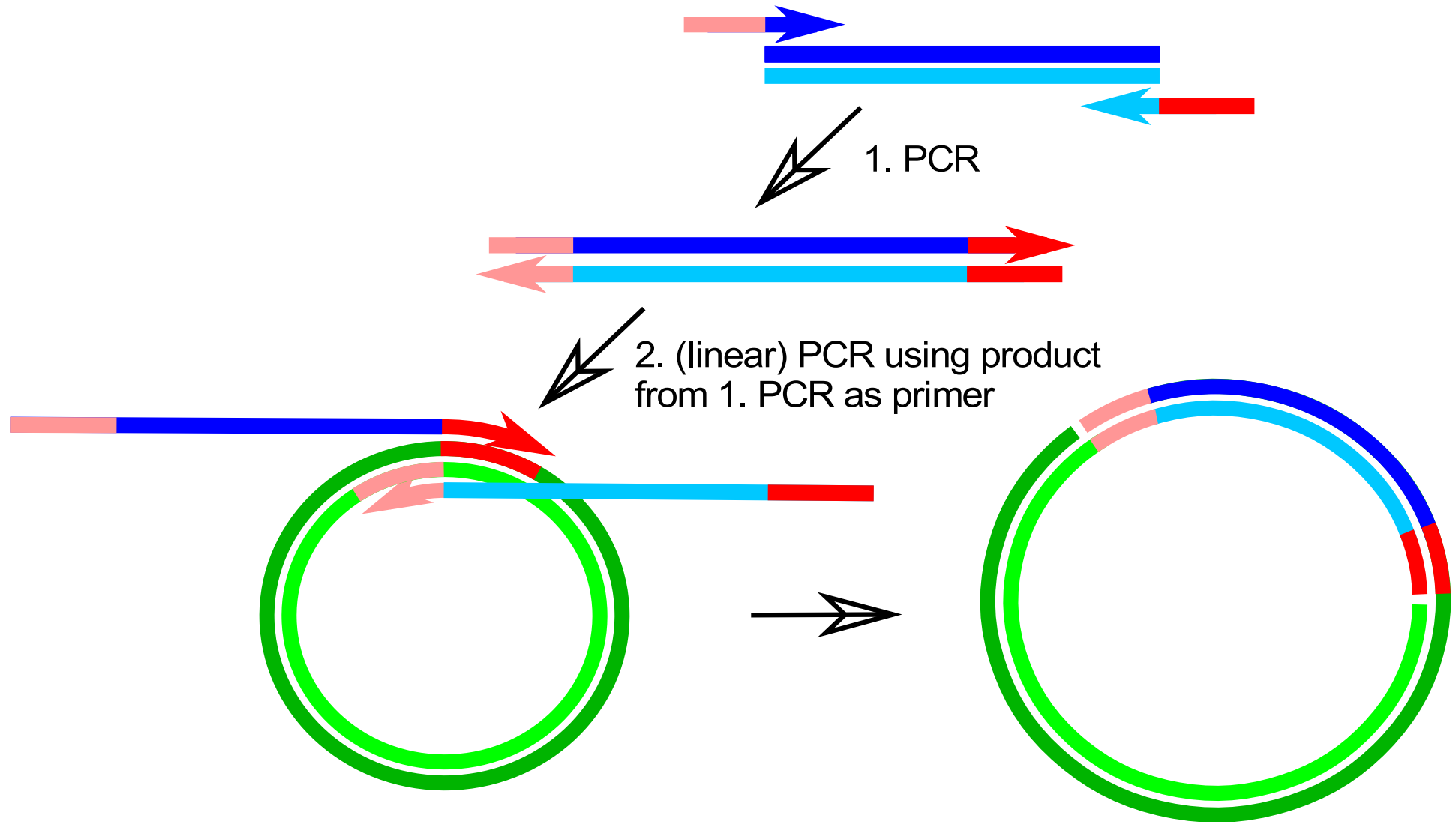


Improved QuikChange



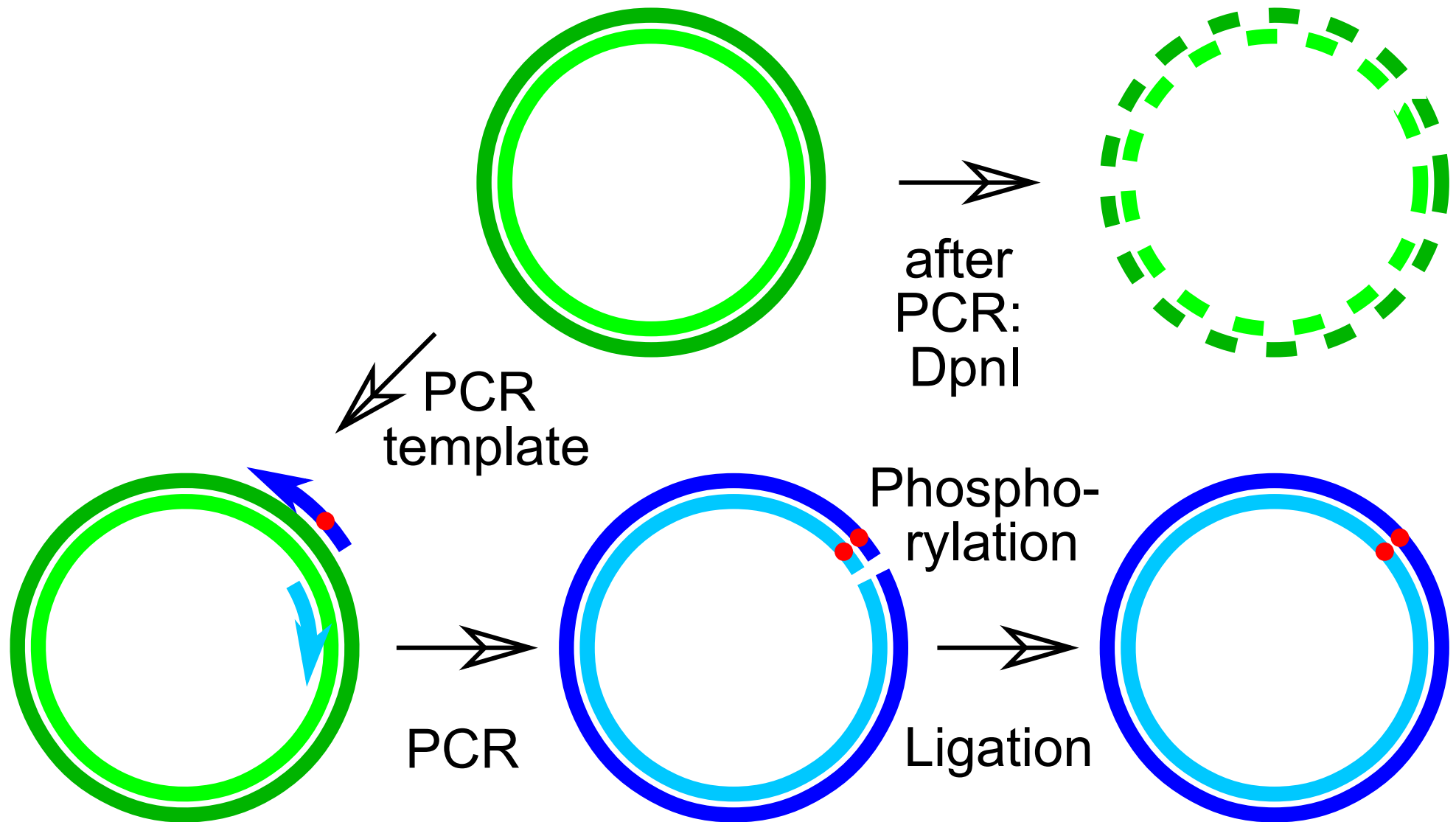


QuikChange Megaprimer (for large insertions)



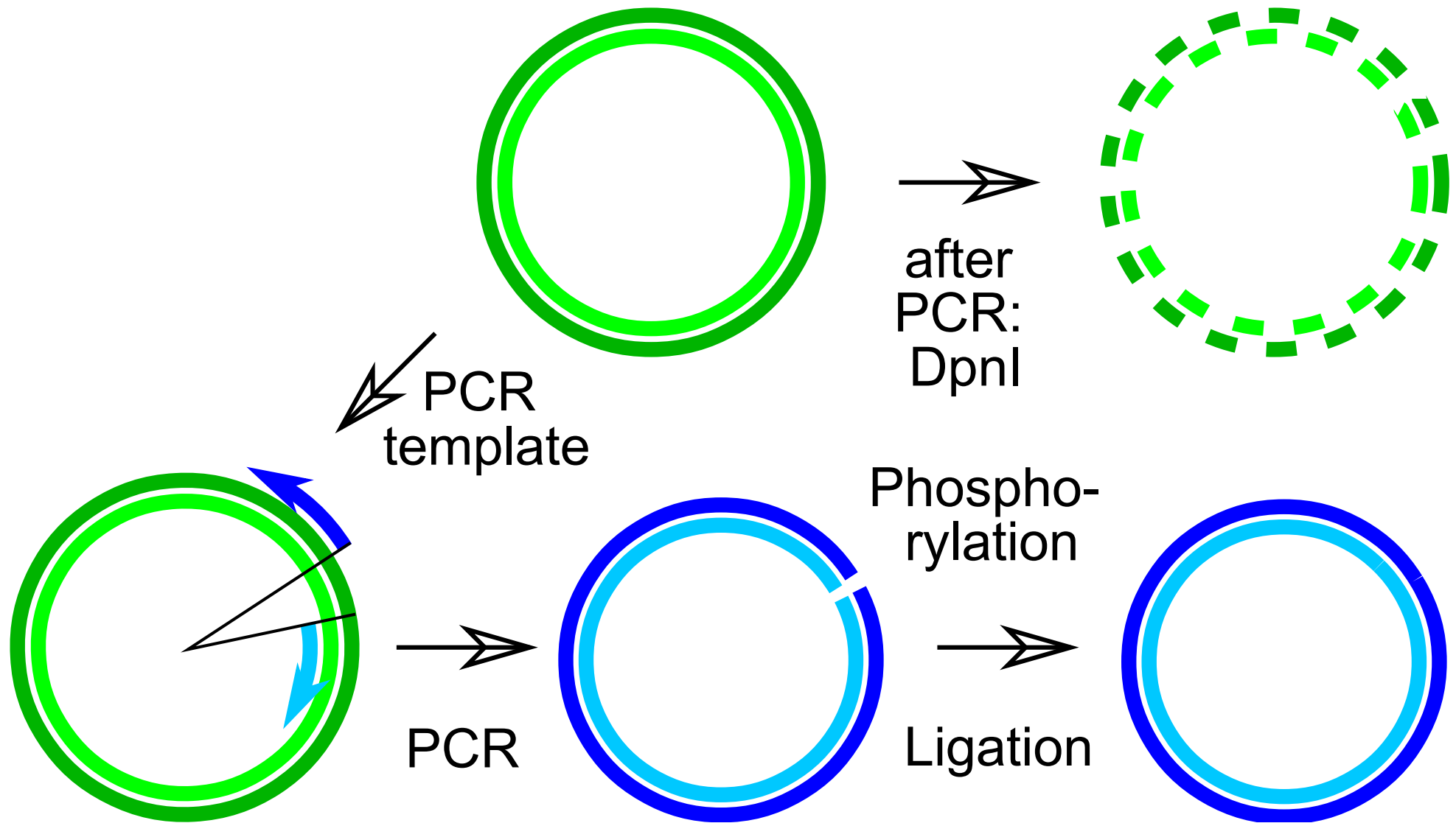


Q5[®]/Phusion[®] SDM: Point mutations



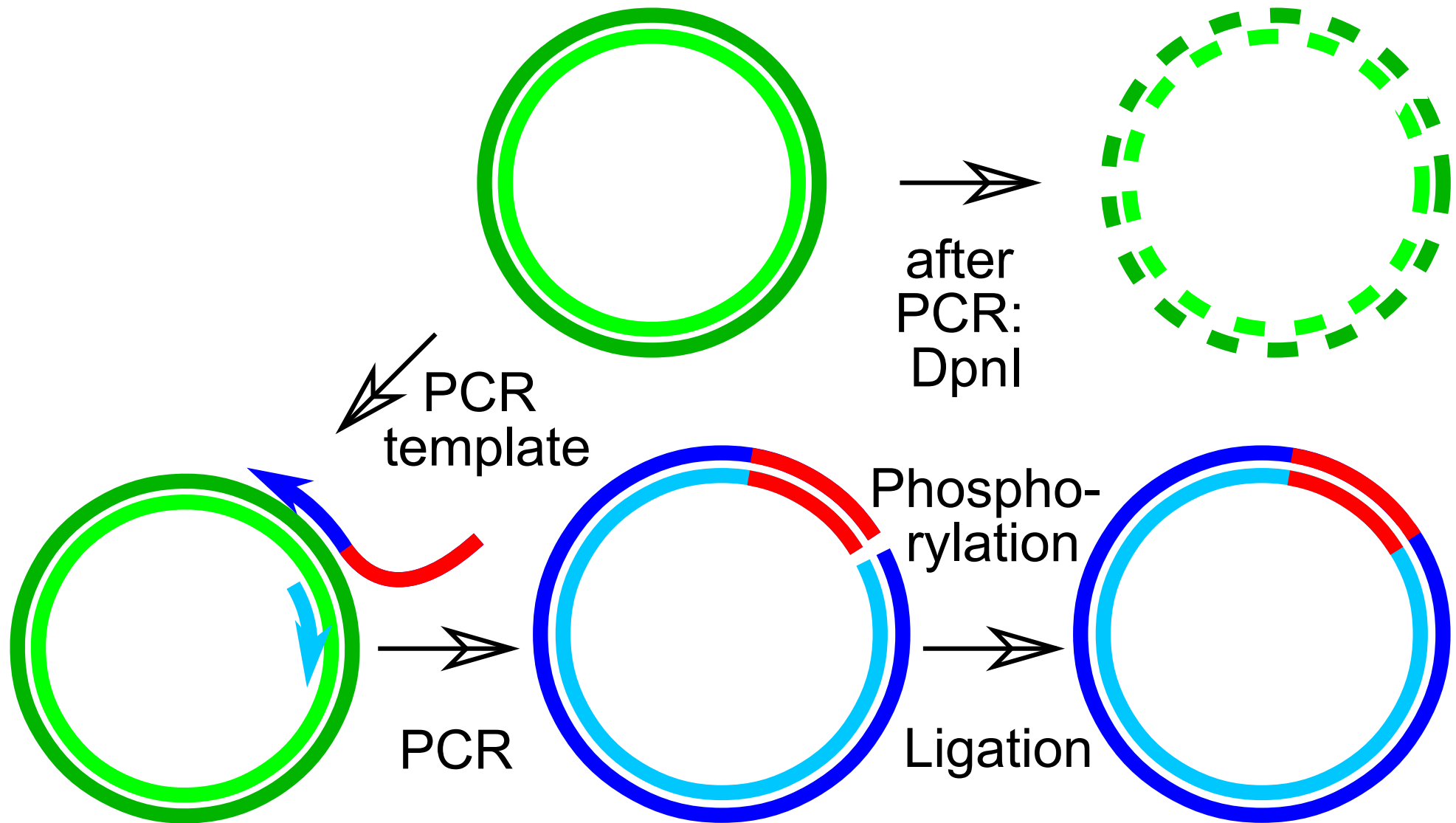


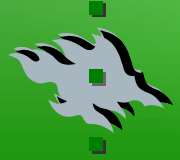
Q5[®]/Phusion[®] SDM: Deletions





Q5[®]/Phusion[®] SDM: Insertions





Whole plasmid mutagenesis: Limitations

- **PCR efficiency**

Primer dimers (solution: separate reactions)

Length of construct (solution: subcloning into minimal vector)

DNA secondary structure (very high/low GC content)

- **Type of mutation**

Large insertions & deletions (solution: Megaprimer)

- **PCR error rate**

All PCR-derived sequences contain potentially mutations and important sequences need to be either sequenced or functionally tested

➔ **Minimize the PCR-derived sequences**



PCR mutagenesis

TTAGTTGGAATTCTAATGAGGATACGGAGATACGGATGTCCGGGTACCAGGATGTC
TTTCAACCTTAAGATTACTCCTATGCCTCTATGCCTACAGGCCCATGGTCCTACAG

AGTTGGAATTCTAATGAGG

original plasmid

TTAGTTGGAATTCTAATGAGGATACGGAGATACGGATGTCCGGGTACCAGGATGTC
TTTCAACCTTAAGATTACTCCTATGCCTCTATGCCTACAGGCCCATGGTCCTACAG

ATGCCTACACCCCATGGTCCTA

mutagenic primer

AGTTGGAATTCTAATGAGGATACGGAGATACGGATGTAGGGTACCAGGAT
TCAACCTTAAGATTACTCCTATGCCTCTATGCCTACATCCCATGGTCCTA

PCR product

AATTCTAATGAGGATACGGAGATACGGATGTAGGGTAC
GATTACTCCTATGCCTCTATGCCTACATCC

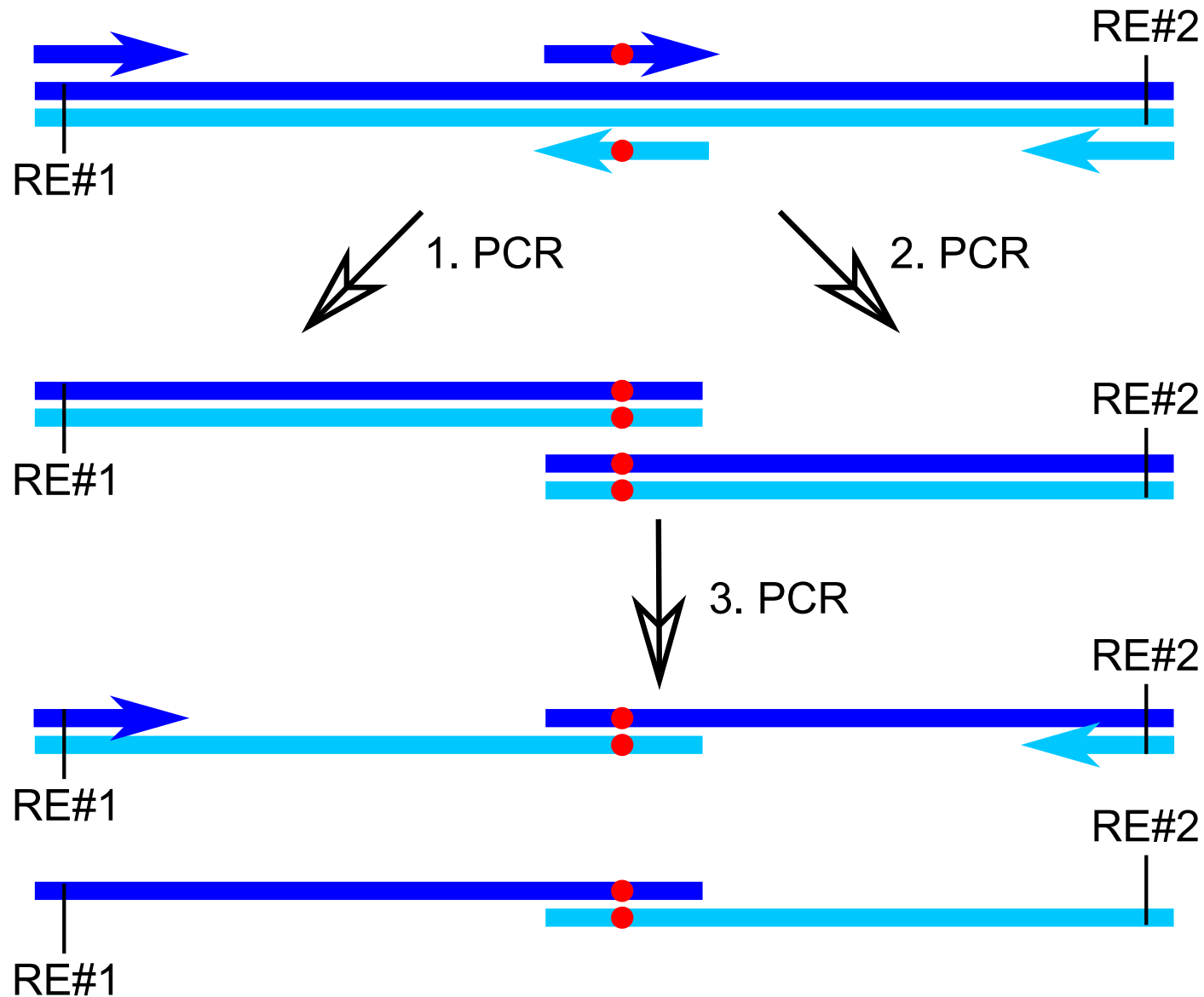
RE-cleaved PCR product

TTAGTTGG
TTTCAACCTTAA

CAGGATGTC
CATGGTCCTACAG

RE-cleaved plasmid

PCR mutagenesis: Overlap extension



SDM by Gibson Assembly

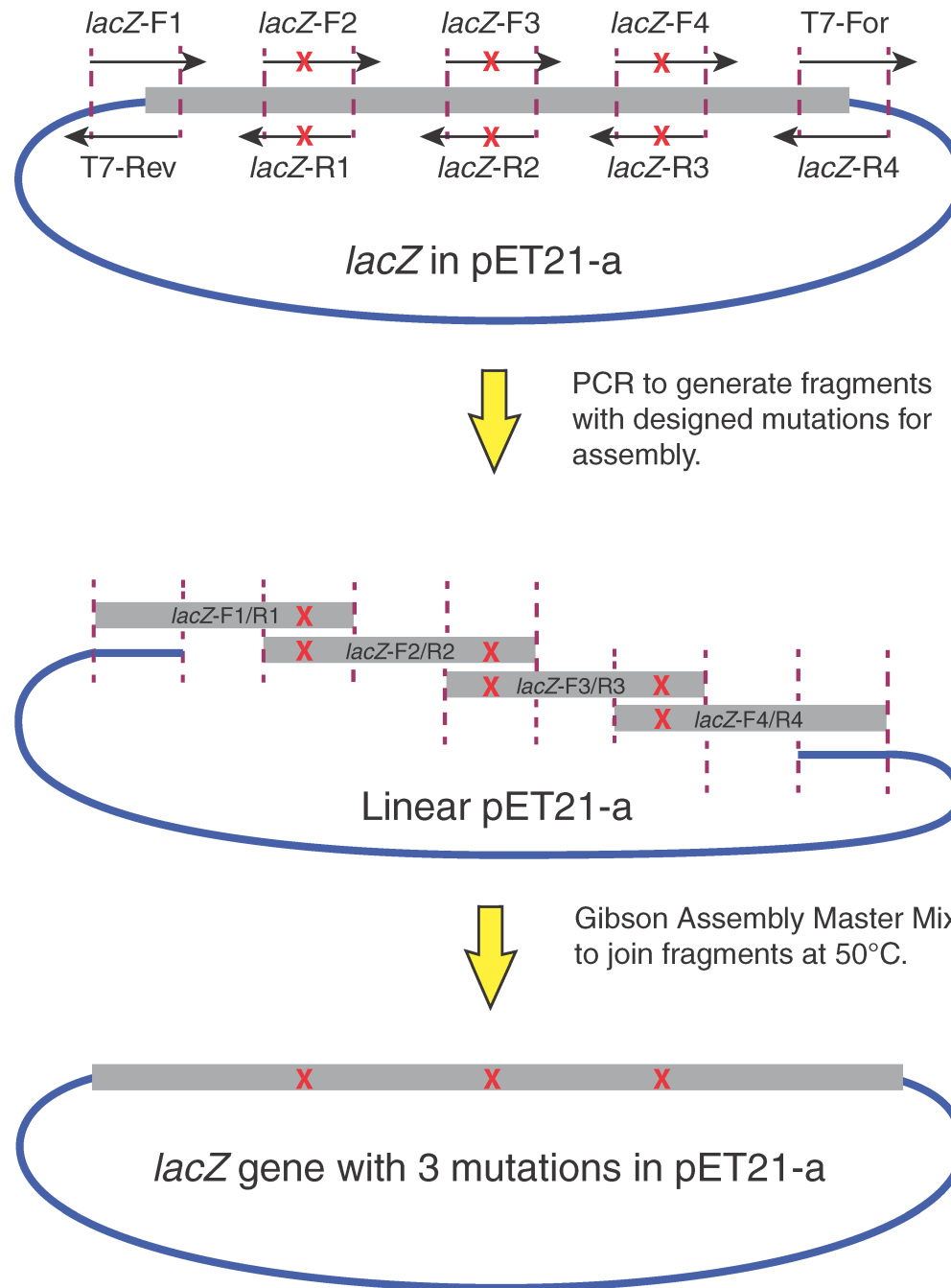


Figure from [NEB](#)

SDM by In-Fusion

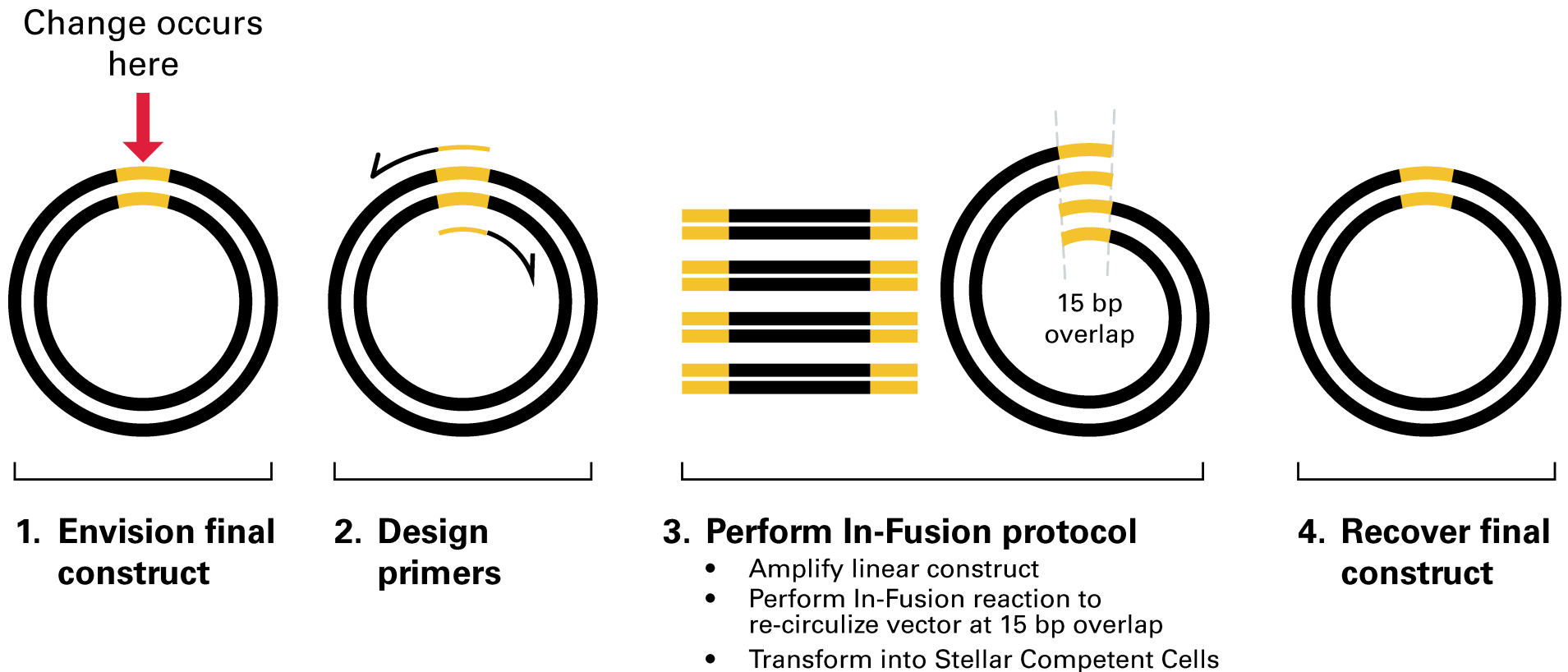


Figure from [Clontech](#)



In-vitro evolution

Methods

Random mutations

- Error-prone PCR (Mn^{2+} or special polymerase)
- Chemical mutagenesis
- Combinatorial cassette mutagenesis
- Mutator strains
- Random Insertion/Deletion (RID)
- Sequence saturation mutagenesis (SeSaM)
- Saturation mutagenesis
- UV irradiation

Homology-based recombination

- DNA shuffling
- DOGS
- Family shuffling
- Family shuffling with restriction enzymes
- RACHITT
- RPR
- StEP
- Genome shuffling

Homology-independent recombination

- Exon shuffling
- DHR
- ITCHY
- THIO-ITCHY
- RM-PCR
- SCRATCHY
- SHIPREC
- SISDC
- YLBS



In-vitro evolution

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Homology-based recombination

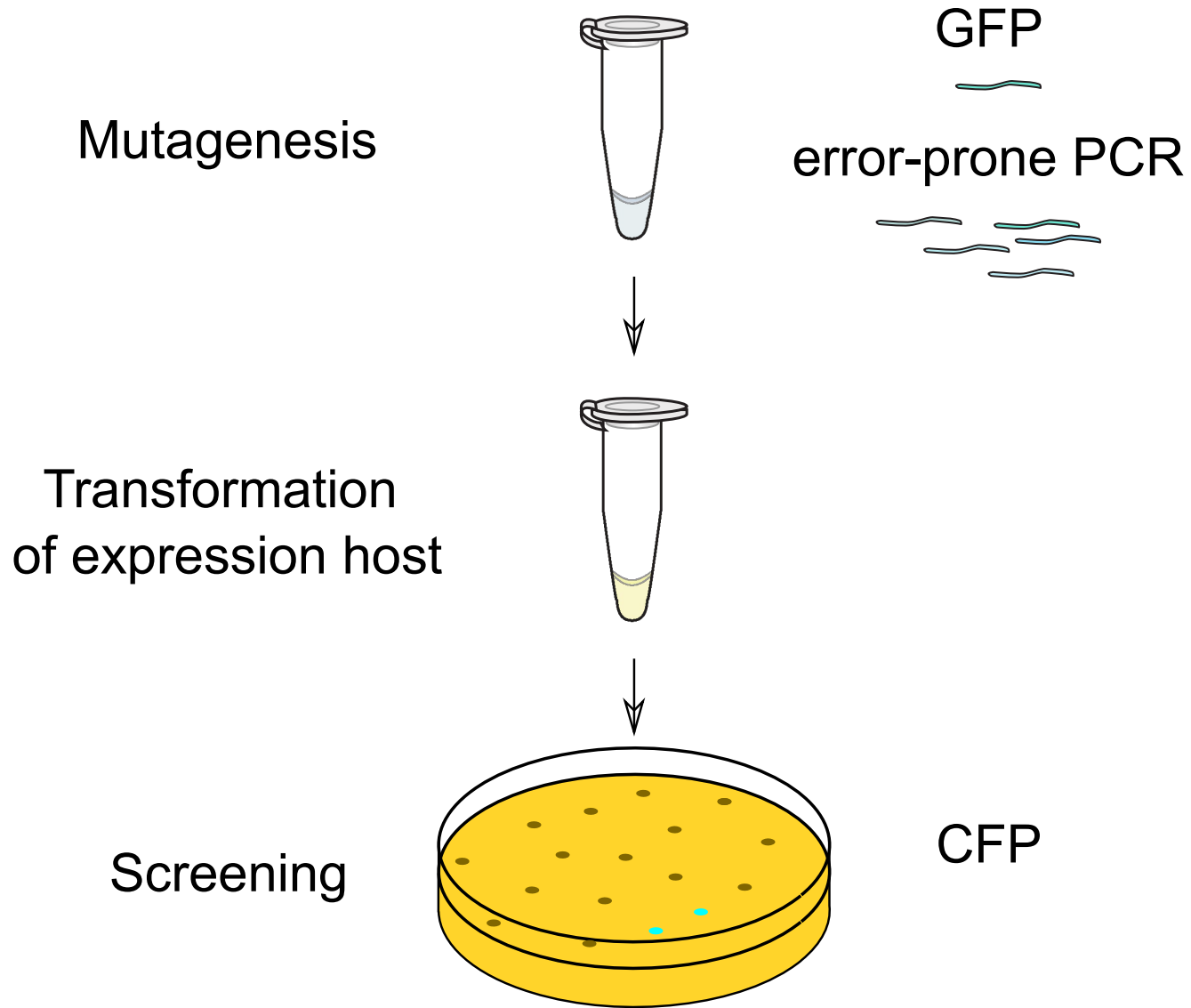
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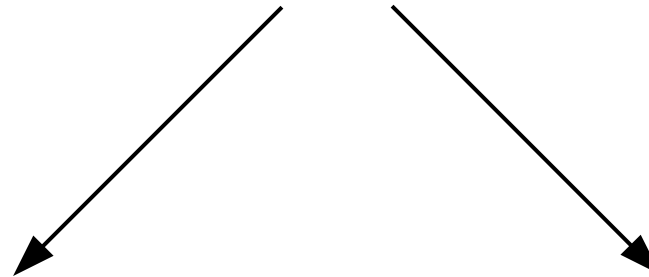
In-vitro evolution





Error-prone PCR

Error-prone PCR



Non-proofreading
polymerase,
 Mn^{2+} ,
dNTP concentration

Mutating PCR enzymes
("Mutazyme")

Polymerase bias, limited diversity (e.g. changing
Tryptophan (TGG) → Isoleucin (AT[TCA]) by
error-prone PCR is almost impossible)

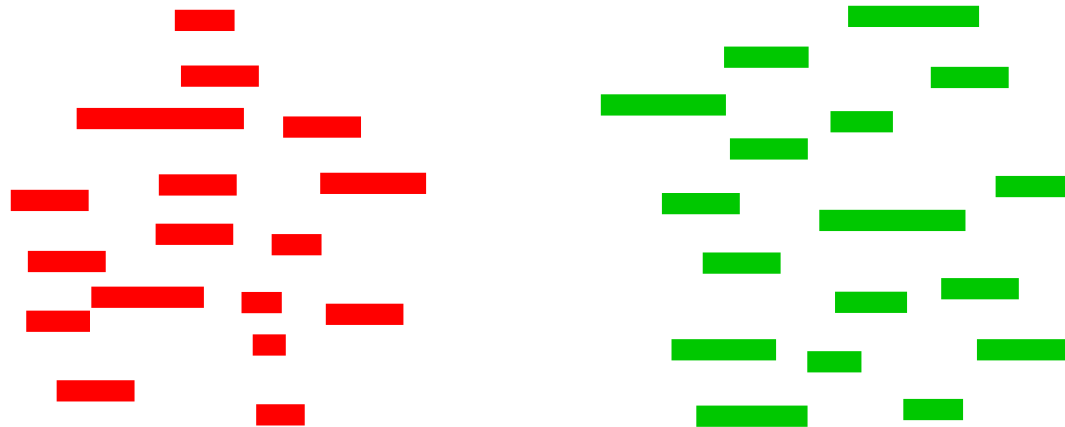


DNA shuffling

Two (or more) homologous sequences



Random fragmentation (DNaseI)

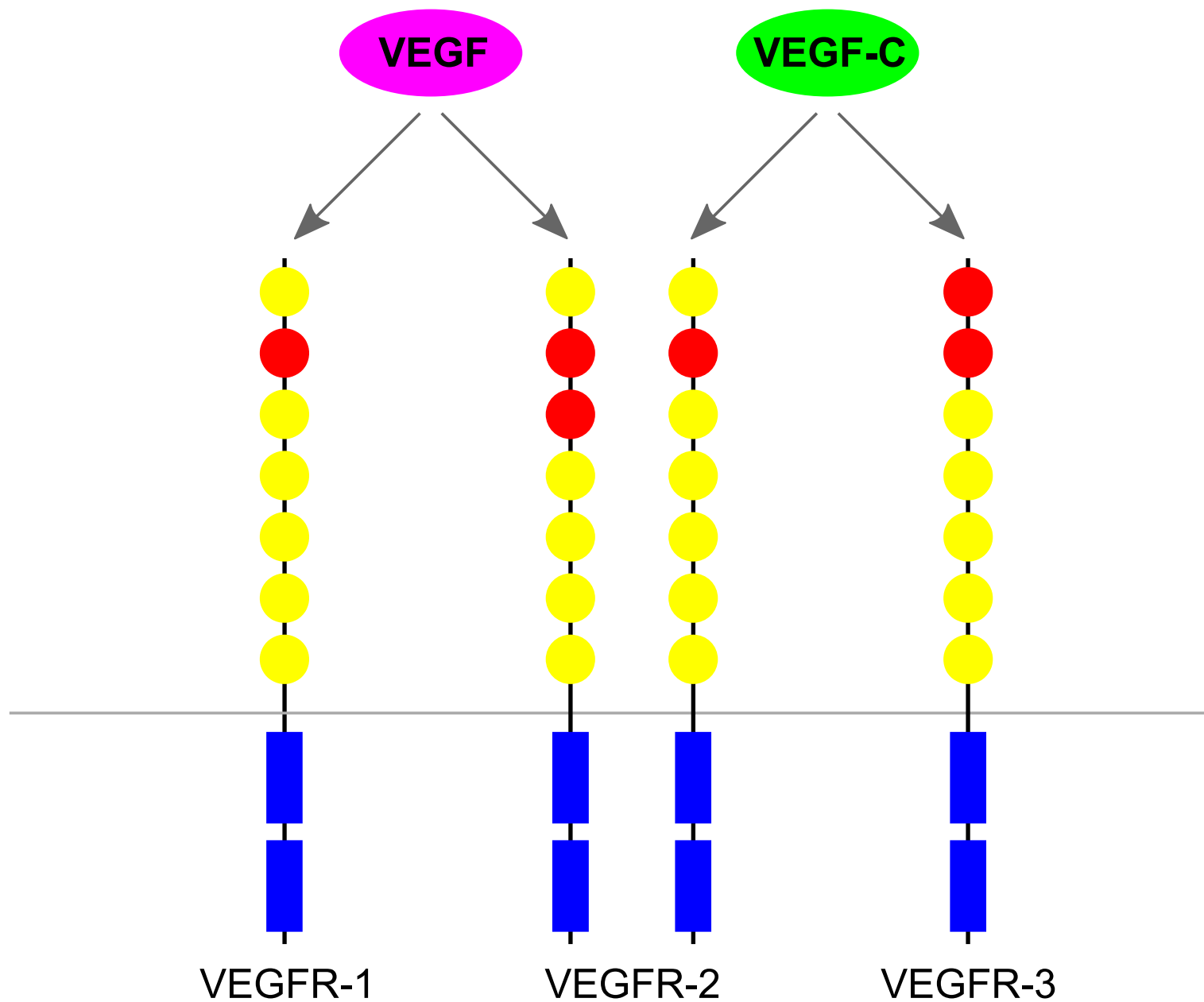


Annealing and repair by PCR



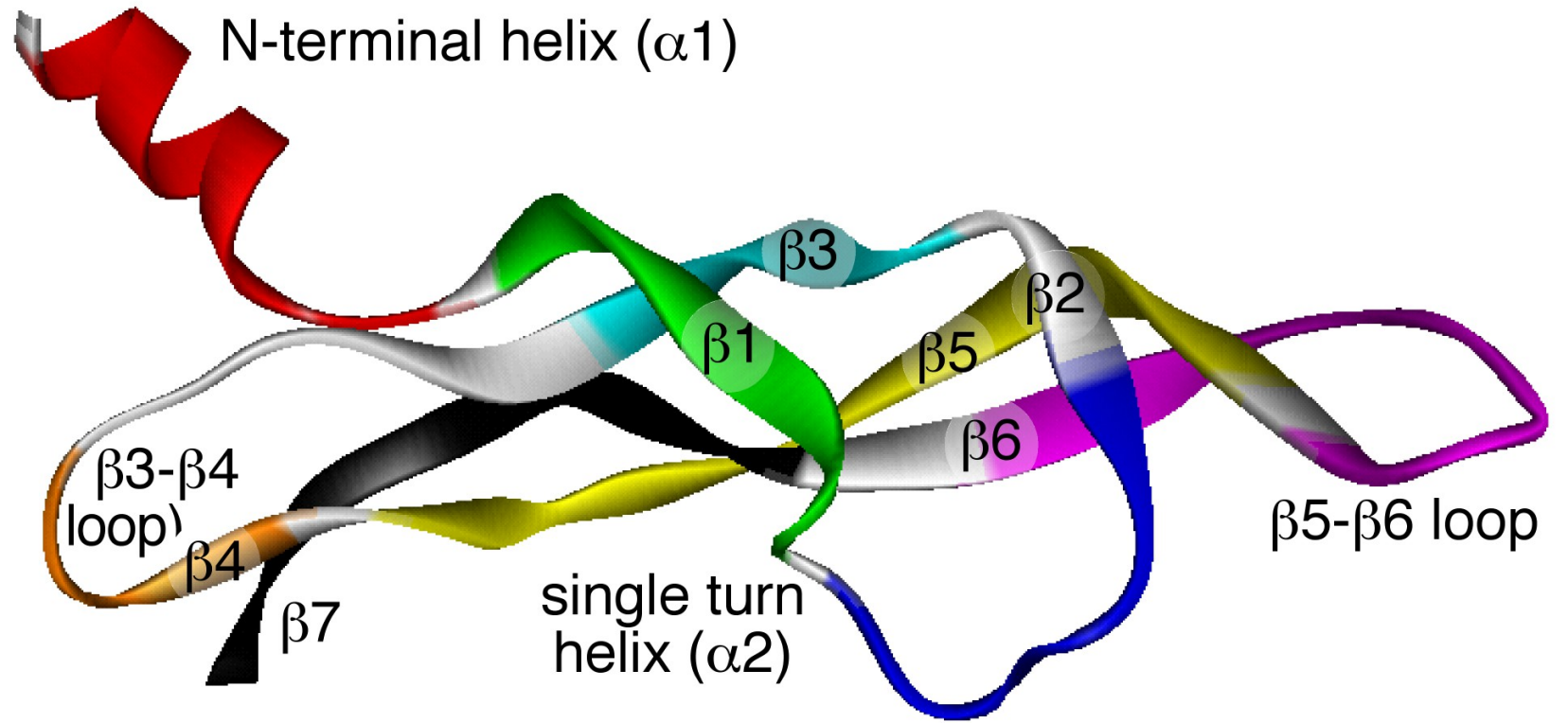


Family shuffling between VEGF-A and VEGF-C





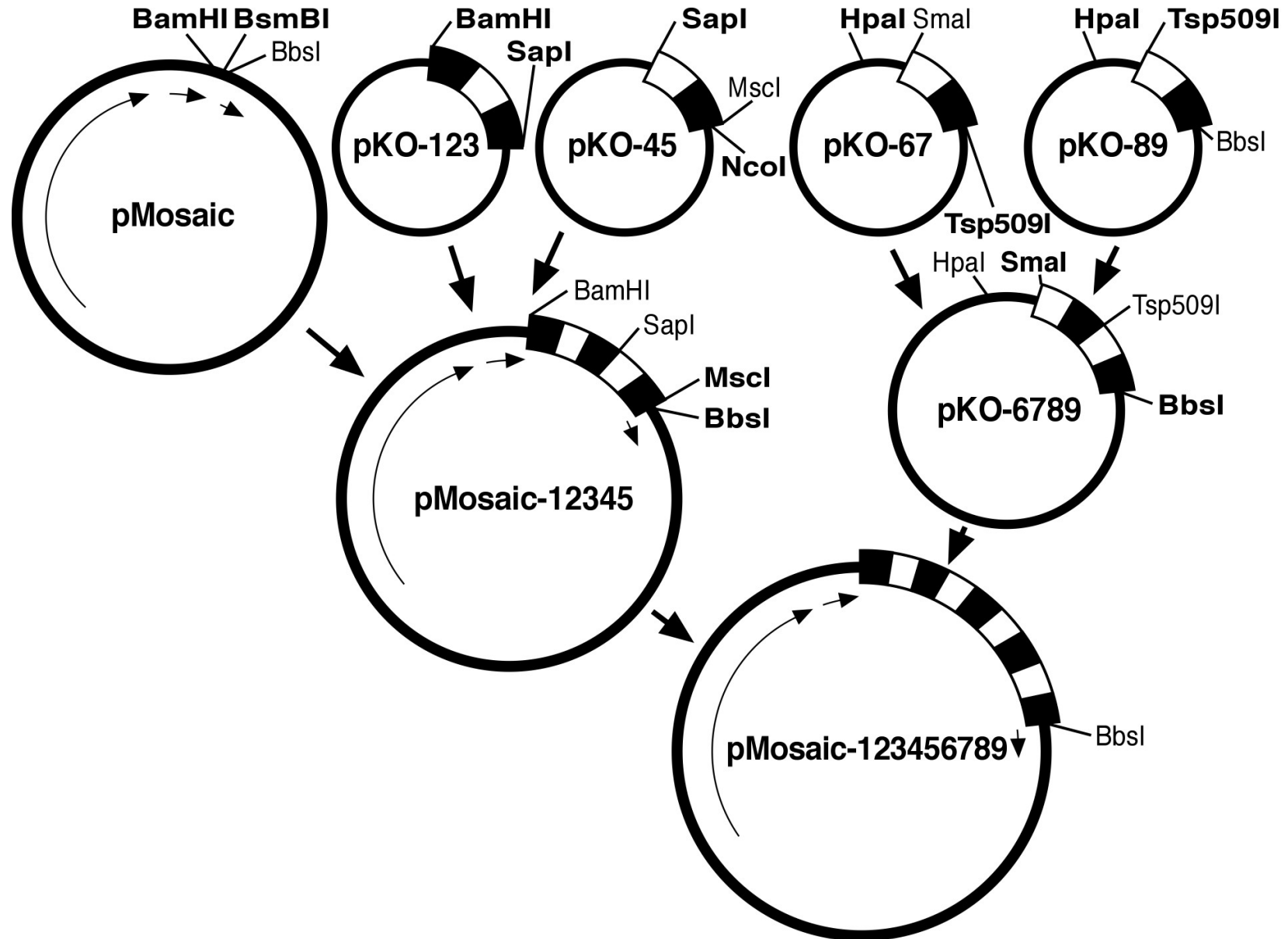
VEGF-A structure



grey	junctions	blue	fragment 4	yellow	fragment 7 ($\beta 5$)
red	fragment 2 ($\alpha 1$)	cyan	fragment 5 ($\beta 3$)	magenta	fragment 8
green	fragment 3 ($\beta 1$)	orange	fragment 5	black	fragment 9 ($\beta 7$)

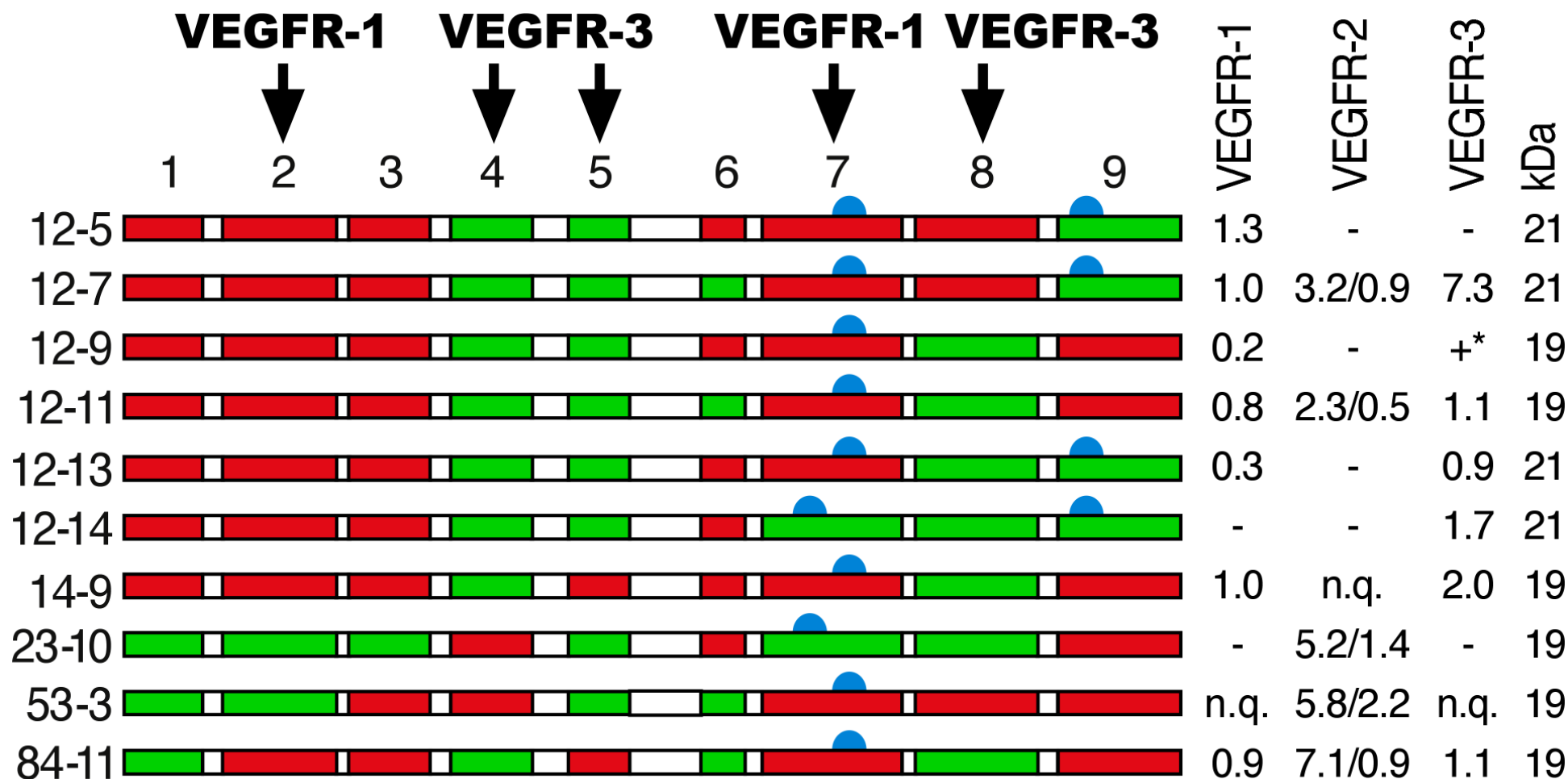


Non-random DNA shuffling





Novel receptor binding profiles in the 2^9 (= 512)-size assorted library





Practical Course

- Participants will be contacted via e-mail
- Participants should form groups of 2 or 4 (if you don't have a group yet, I will assign you into one)
- Meetings with all groups within the next two weeks to discuss, plan and prepare the projects